DETERMINATION OF LONG DISTANCE TRANSPORT OF Cs⁺, Co²⁺ AND Zn²⁺ IONS IN VASCULAR PLANTS BY AUTORADIOGRAPHY AND GAMMA-SPECTROMETRY

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Abstract: Heavy metals and radionuclides can enter the food chain via cereals and vegetables grown in contaminated soils. In the case of microelements such as zinc, studies have not focused only to assessing its environmental risk, but also to enhancing its uptake by plants as an important growth-limiting factor. In our study, digitalized autoradiograms of whole plants of celery (Apium graveolens L.), tobacco (Nicotiana tabacum L.) and sunflower (Helianthus annuus L.) grown in hydroponic nutrient media spiked with 137CsCl, ⁶⁰CoCl₂ and ⁶⁵ZnCl₂ were used for quantitative determination of uptake, long-distance transport and distribution of Cs⁺, Co²⁺ and Zn²⁺ ions in plant tissues. Results from autoradiography and gammaspectrometry of plants showed, that cesium was translocated to aboveground part of the plants with the shoot-to-root ratio 1.0: 0.6. On the contrary, cobalt and zinc were more immobilized in roots, with the shoot-to-root ratio up to 1.0:3.8. The highest concentration of cesium, cobalt and zinc, expressed in specific radioactivity per unit of leaf surface (Bq/cm2) was found in top, rapidly growing leaves, the lowest concentration in the oldest leaves in low position. Detection limits 3, 2 and 14 Bq/cm² by using X-ray film for ¹³⁷Cs, ⁶⁰Co and ⁶⁵Zn, respectively were obtained. These data correspond to detection limits 10.5 pg Cs⁺/cm², 7.2 pg Co²⁺/cm² and 785 pg Zn²⁺/cm² at specific radioactivity of commercially available ¹³⁷CsCl, ⁶⁰CoCl₂ and ⁶⁵ZnCl₂. Resolutions 1-2 mm was sufficient for visualization of metal uptake and distribution in roots, stalks, leaves and leaf venation. Obtained data are part of quantitative study of uptake and translocation of both low level-radioactive contamination in plants and microelements applied as fertilizers.

Keywords: cobalt, cesium, zinc, distribution, vascular plants, autoradiography

1. Introduction

Heavy metals are present in soils and aqueous streams as both natural components or as a result of human activity i.e. metal-rich mine tailings, metal smelting, electroplanting, energy and fuel production, intensive agriculture, etc. (RASKIN *et al.*, 1994). Radionuclides exist in the environment either naturally or artificially by aboveground nuclear testing, nuclear accidents and nuclear power generation (ZHU and SHAW, 2000).

Cesium 137 Cs ($\tau = 30.17$ y) is produced during the fission of actinides or the neutron bombardment of 133 Cs or 136 Ba (WHITE and BROADLEY, 2000). The behavior of Cs in soils resembles that of K and, even in soils with high clay content, Cs is available for uptake by plants since Cs-fixation tends towards a reversible steady-state.

Cobalt usually occurs in the environment in association with other metals such as copper, nickel, manganese and arsenic. Background soil cobalt levels range from 1.6

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to 21.5 mg.kg⁻¹ (PEREZ-ESPINOZA *et al.*, 2005). Cobalt 60 Co (τ = 5.27 y) is also present in low-level radioactive wastes.

Although zinc is an essential nutrient as a trace element for animals, plants, and microorganisms, it is toxic to these organisms when present at supranormal concentrations. Zinc is a trace element essential for cell proliferation and differentiation. It is a structural constituent of many enzymes and proteins, including metabolic enzymes, transcription factors, and cellular signaling proteins.

There is considerable interest in remediation of sites contaminated by these elements using extraction by plants that do not enter the human food chain. Several comprehensive reviews have been written, summarizing many important aspects of this plant-based technology (LASAT, 2002; DUSHENKOV, 2003; SURESH and RAVISHANKAR, 2004; PILON-SMITS, 2005; LEDUC and TERRY, 2005; MARMIROLI *et al.*, 2006).

The ecological problems related to heavy metals and radionuclides are not dependent only on their total content and radioactivity in the soil, but rather on their form of bonding and therefore their bioavailability. Therefore, several studies have been conducted using seedlings or adult plant, which have been cultivated in hydroponic conditions (see e.g. PAGE and FELLER, 2005; SOUDEK *et al.*, 2006).

In our study, digitalization of autoradiograms of whole plants of celery (*Apium graveolens* L.), tobacco (*Nicotiana tabacum* L.) and sunflower (*Helianthus annuus* L.) after cultivation in hydroponic nutrient media spiked with ¹³⁷CsCl, ⁶⁰CoCl₂ and ⁶⁵ZnCl₂ was used as the method for quantitative determination of uptake, long-distance transport and distribution of Cs⁺, Co²⁺ and Zn²⁺ ions in the plant tissues.

2. Materials and methods

2.1 Plant material

Seeds of celery (*A. graveolens* L.), tobacco (*N. tabacum* L.) and sunflower (*H. annuus* L.) were germinated and grown in pots filled with granulated perlite as an inert carrier and watered with diluted hydroponic medium according to HOAGLAND (1920) in day/night period 12/12 h (illumination with 2 tubes Brilliant daylight - 6000 K, 1 300 lm and Tropic sun – 4 700 K, 1 000 lm, SERA) at 22±2°C. After 5 weeks (for celery and sunflower) or 8 weeks (for tobacco), seedlings were gently removed from perlite and roots were washed free of any adhering perlite fragments by distilled water. Plants were pre-cultivated for one week in vessels with a cover to protect plant roots against lights in 25 % Hoagland medium without perlite support.

2.2 Bioaccumulation experiments

For experiments selected plants were cultivated at $22\pm2^{\circ}$ C in 25 % Hoagland medium supplemented with 10 µmol.L⁻¹ CsCl, 10 µmol.L⁻¹ CoCl₂ or 5 µmol.L⁻¹ ZnCl₂ and spiked with ¹³⁷Cs, ⁶⁰Co or ⁶⁵Zn at illumination 2 000 lx with 12/12 h day/night cycle. In time intervals aliquot samples of cultivation media were measured for

determination of remaining radioactivity by gamma-spectrometry. The amounts of transpired water were determined by measuring the weight changes of cultivation vessels. The amount of water lost by plant transpiration was replenished with fresh 25 % Hoagland medium to initial solution volume. At the end of experiments plants were harvested and roots carefully rinsed in distilled water. In roots, stalks and leaves incorporated radioactivity was measured by gamma-spectrometry and topography visualized by autoradiography as well.

2.3 Autoradiography

Plants were pressed between two filter papers and air-dried for 7 days at 20°C. ¹³⁷Cs, ⁶⁰Co and ⁶⁵Zn in dried plant were detected by autoradiography by exposing X-ray films (HR-GB 100 NIF, FUJIFILM, JP) for 40 days at 20°C. Developed films were scanned and data were converted to colour scale gradient by software Photoshop CS2 ver. 9.0 (ADOBE, USA).

2.4 Radiometric analysis

Gamma-spectrometric scintillation detector 54BP54/2-X with well type crystal NaI(Tl) (Scionix, NL) and data processing software Scintivision32 (ORTEC, USA) were used for ¹³⁷Cs, ⁶⁰Co and ⁶⁵Zn determination in plant and solution. Counting time 600 s allowed obtaining data with measurement error <2 %, which do not reflect other source of errors. Standardized ¹³⁷CsCl (5.723 MBq.mL⁻¹; 20 mg.L⁻¹ CsCl + 3 g.L⁻¹ HCl), ⁶⁰CoCl₂ (5.571 MBq.mL⁻¹; 20 mg.L⁻¹ CoCl₂ + 3 g.L⁻¹ HCl) and ⁶⁵ZnCl₂ (0.892 MBq.mL⁻¹; 50 mg.L⁻¹ ZnCl₂ + 3 g.L⁻¹ HCl) were obtained from CMI (CZ).

3. Results and discussion

Kinetics of cesium and cobalt bioaccumulation by celery (*A. graveolens* L.), tobacco (*N. tabacum* L.) and sunflower (*H. annuus* L.) hydroponically growing in nutrient media spiked with ¹³⁷CsCl and ⁶⁰CoCl₂ were described in our previous papers (HORNIK *et al.*, 2005; BARATOVA *et al.*, 2006; VRTOCH *et al.*, 2006; VRTOCH *et al.*, 2007). We found, that bioaccumulation of ¹³⁷Cs and ⁶⁰Co by these plants was proportional to the transpiration rate.

The aim of this paper is to characterize long-distance transport of $^{137}\text{Cs}^+$, $^{60}\text{Co}^{2+}$ and $^{65}\text{Zn}^{2+}$ in celery, tobacco and sunflower.

Radiometric analysis showed, that cesium accumulated by celery and tobacco from nutrient solution containing sub-inhibitory concentrations $10 \,\mu \text{mol.L}^{-1}$ and $2.4 \,\mu \text{mol.L}^{-1}$ CsCl was transported from roots to shoots. The highest cesium concentration was found in top leaves. (Fig. 1, Fig. 2). Similar pattern of cesium transport in sunflower was observed in our previous paper (HORNIK *et al.*, 2005). Shoot-to-root concentration ratio was within the range 1: 0.60 - 1: 0.88 (Tab. 1). Concentration of cesium expressed in specific radioactivity per unit of leaf surface (Bq/cm²) was 119-

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times higher in top, rapidly growing celery leaves than in the oldest celery leaves and 12-times higher in top tobacco leaves (Fig. 3).

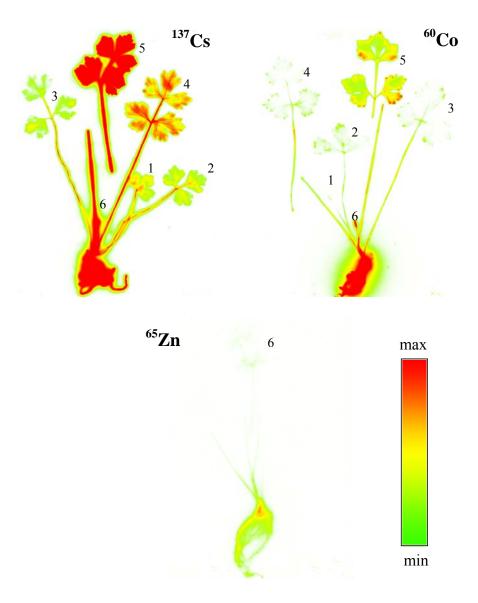


Fig. 1: Autoradiographic visualization of 137 Cs, 65 Co and 65 Zn distribution in celery (*A. graveolens* L.) after 8 days cultivation in 25 % Hoagland medium. Initial concentrations $C_0 = 10 \, \mu \text{mol.L}^{-1} \, \text{CsCl}$, $10 \, \mu \text{mol.L}^{-1} \, \text{CoCl}_2$ or 5 $\mu \text{mol.L}^{-1} \, \text{ZnCl}_2$, spiked with 167.6 kBq.L⁻¹ ¹³⁷CsCl, 91 kBq.L⁻¹ ⁶⁰CoCl₂ or 68 kBq.L⁻¹ ⁶⁵ZnCl₂. Uptake: Cs – 37 %; Co – 97 %; Zn – 40 %.

Numbering of leaves in the order: 1. oldest leaves, 6. youngest leaves.

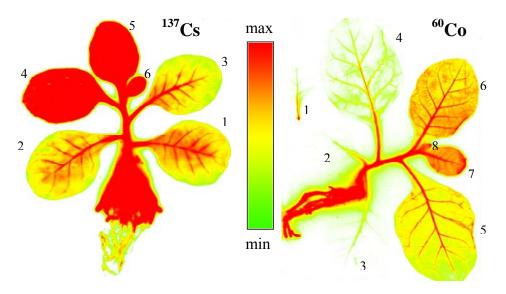


Fig. 2: Autoradiographic visualization of ^{137}Cs and ^{65}Co distribution in tobacco (*N. tabacum* L.) after 9 days cultivation in 25 % Hoagland medium. Initial concentrations $C_0=2.4~\mu mol.L^{-1}~CsCl$ or 25 $\mu mol.L^{-1}~CoCl_2,$ spiked with 283 kBq.L $^{-1}$ $^{137}CsCl$ or 29.7 kBq.L $^{-1}$ $^{60}CoCl_2.$ Uptake: Cs-65%; Co-65%. Numbering of leaves in the order: 1. oldest leaves, 8. youngest leaves.

Long-distance transport of both cobalt and zinc in celery, to bacco and sunflower cultivated in nutrient media in sub-inhibitory concentration $0.385-25~\mu mol.L^{-1}~CoCl_2$ or $5~\mu mol.L^{-1}~ZnCl_2$ was significantly lower than in the case of cesium. Cobalt and zinc were mainly trapped in roots. Shoot-to-root concentration ratio up to 1:3.78 was observed (Tab. 1). Concentration of cobalt and zinc, expressed in specific radioactivity per unit of leaf surface (Bq/cm²) of celery, to bacco and sunflower was approx. 50-times higher in top, younger leaves than in the oldest leaves (Fig. 3).

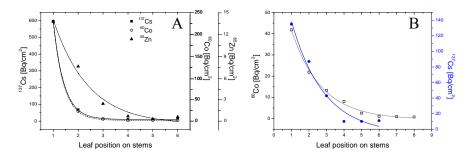


Fig. 3: Specific radioactivity of ¹³⁷Cs, ⁶⁰Co and ⁷⁵Zn accumulated in leaves of celery (A) and tobacco (B), calculated for leaf surface (Bq/cm²). Numbering of leaves from top to root.

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Mobility of monovalent cesium in tobacco, celery and sunflower plants is higher than mobility of bivalent zinc and cobalt. This fact is in an agreement with well described transport mechanisms of cesium, comparable with transport mechanisms of essential potassium K⁺ ions (WHITE and BROADLEY, 2000). Cesium, with the exception of some synthetic compounds such as crown ethers, does not form stable organic complexes (DOZOL et al., 2004).

On the other hand, cobalt and zinc can react with low and high molecular organic components of plants (OWEN et al., 2002). SALT et al. (1999) found, that Zn was mostly complexed to histidine in roots, transported as Zn²⁺ in the xylem sap, and complexed to organic acids in leaves. In dependence on pH value, zinc and cobalt can form charged species by the reaction with CO₂ and OH ions (DANG et al., 1996; GÜNTHER and KASTENHOLZ, 2005).

PAGE and FELLER (2005) showed that zinc was translocated from root to young wheat leaves within 50 days nearly uniformly in all leaves of the shoot. On the contrary, prevailing part of cobalt even after 50 days remained in root and transport to all leaves including apex leaves was negligible.

Mobility of zinc and cobalt in vascular plants within time intervals, comparable with duration of vegetation period, will be the topic of our next study.

Tab. 1: Distribution of cesium, cobalt and zinc in shoots and roots of celery (A. graveolens L.), tobacco (N. tabacum L.) and sunflower (H. annuus L.) cultivated in 25 % Hoagland medium spiked with 137CsCl 60CoCl2, or 65ZnCl2.

Nuclide -	Shoot-to-root ratio		
	Celery ¹	Tobacco ²	Sunflower ³
¹³⁷ Cs	1.0:0.60	1.0:0.88	1.0:0.64
⁶⁰ Co	1.0:3.78	1.0:1.02	1.0 : 2.03
⁶⁵ Zn	1.0:1.29	-	_

^{1 -} After 8 day cultivation, C₀ = 10 μmol.L⁻¹ CsCl (168 kBq.L⁻¹), C₀ = 10 μmol.L⁻¹ CoCl₂ (91 kBq.L⁻¹) or $C_0 = 5 \mu \text{mol.L}^{-1} \text{ZnCl}_2 (68 \text{ kBq.L}^{-1})$

Detection limits obtained by autoradiography of the whole plants was 3, 2 and 14 Bq/cm² for ¹³⁷Cs, ⁶⁰Co and ⁶⁵Zn, respectively. It corresponds to detection limits 10.5 pg Cs/cm², 7.2 pg Co/cm² and 785 pg Zn/cm² at specific radioactivities of commercially available ¹³⁷CsCl, ⁶⁰CoCl₂ and ⁶⁵ZnCl₂. Resolution 1-2 mm was sufficient for visualization of distribution in roots, stalks, leaves and leaf venation.

4. Conclusions

Autoradiography and gamma-spectrometry of celery (Apium graveolens L.), tobacco (Nicotiana tabacum L.) and sunflower (Helianthus annuus L.) after

^{2 -} After 9 day cultivation, $C_0 = 2.4 \ \mu mol.L^{-1} \ CsCl \ (283 \ kBq.L^{-1})$ or $C_0 = 25 \ \mu mol.L^{-1} \ CoCl_2 \ (29.7 \ kBq.L^{-1})$ 3 - After 9 day cultivation, $C_0 = 1.25 \ \mu mol.L^{-1} \ CsCl \ (25 \ kBq.L^{-1})$ or $C_0 = 0.385 \ \mu mol.L^{-1} \ CoCl_2 \ (25 \ kBq.L^{-1})$. See also HORNIK et al., (2005).

cultivation in hydroponic nutrient media spiked with ¹³⁷CsCl, ⁶⁰CoCl₂ and ⁶⁵ZnCl₂ was used for quantitative determination of uptake and long-distance transport of Cs⁺, Co²⁺ and Zn²⁺ ions in the plant tissues.

Cesium was traslocated to the aboveground part of the plants with the shoot-to-root ratio 1.0: 0.6. On the contrary, cobalt and zinc was more immobilized in roots, with the shoot-to-root ratio up to 1.0: 3.8.

The highest concentration of cesium, cobalt and zinc expressed in specific radioactivity per unit of leaf surface (Bq/cm²) was found in top, rapidly growing leaves, the lowest concentration in the oldest leaves in low position.

Detection limits 3, 2 and 14 Bq/cm² by using X-ray film for ¹³⁷Cs, ⁶⁰Co and ⁶⁵Zn, respectively were obtained. It corresponds to detection limits 10.5 pg Cs⁺/cm², 7.2 pg Co²⁺/cm² and 785 pg Zn²⁺/cm² at specific radioactivity of commercially available ¹³⁷CsCl, ⁶⁰CoCl₂ and ⁶⁵ZnCl₂. Resolution 1-2 mm was sufficient for visualization of distribution in roots, stalks, leaves and leaf venation.

Obtained data are part of quantitative study of uptake and translocation of both low-level radioactive contamination in plants and microelements applied as fertilizers.

References

- BARATOVA, Z., SEKACOVA, J., HORNIK, M., PIPISKA, M., AUGUSTIN, J.: Bioaccumulation of ¹³⁷Cs and ⁶⁰Co by mustard *Sinapis alba* L. and celery *Apium graveolens* L.. Nova Biotechnol., VI-I, 2006, 7-18.
- DANG, Y.P., TILLER, K.G., DALAL, R.C., EDWARDS, D.G.: Zinc speciation in soil solutions of vertisols. Aust. J. Soil Res., 34, 1996, 369-383.
- DOZOL, J.F., DOZOL, M., MACIAS, R.M.: Extraction of strontium and cesium by dicarbollides, crown ethers and functionalized calixarenes. J. Incl. Phenom. Macrocyclic Chem., 38, 2000, 1-22.
- DUSHENKOV, S.: Trends in phytoremediation of radionuclides. Plant Soil, 249, 2003, 167-175.
- GÜNTHER, K., KASTENHOLZ, B.: Speciation of zinc. *In:* R. CORNELIS; H. CREWS; J. CARUSO; K.G. HEUMANN (Eds.) Handbook of elemental speciation II: Species in the environment, food, medicine and occupational health. John Wiley & Son, New York, 2005, 488-506.
- HOAGLAND, D.R.: Optimum nutrient solutions for plants. Science, 52, 1920, 562-564
- HORNIK, M., PIPISKA, M., VRTOCH, L., AUGUSTIN, J., LESNY, J.: Bioaccumulation of ¹³⁷Cs and ⁶⁰Co by *Helianthus annuus*. Nukleonika, 50(1), 2005, S49-S52.
- LASAT, M.M.: Phytoextraction of toxic metals A review of biological mechanisms. J. Environ. Qual., 31, 2002, 109-120.
- LEDUC, D.L., TERRY, N.: Phytoremediation of toxic trace elements in soil and water. J. Ind. Microbiol. Biotechnol., 32, 2005, 514-520.
- MARMIROLI, N., MARMIROLI, M., MAESTRI, E.: Phytoremediation and phytotechnologies: a review for the present and future. *In:* I. TWARDOWSKA, ALLEN, H.E., HÄGGBLOM, M.M., STEFANIAK, S. (Eds.) Soil and water

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pollution monitoring, protection and remediation, Springer, Dordrecht, The Netherlands, 2006, 403-415.

- OWEN, M., GRILL, E., GOLAN-GOLGHIRSH, A., KUTCHAN, T.M., ZENK, M.H.: Increase of free cysteine and citric acid in plant cells exposed to cobalt ions. Phytochem., 60, 2002, 467-474.
- PAGE, V., FELLER, U.: Selective transport of zinc, manganese, nickel, cobalt and cadmium in the root system and transfer to the leaves in young wheat plants. Ann. Bot., 96, 2005, 425-434.
- PEREZ-ESPINOZA, A., MORAL, R., MORENO-CASELLES, J., CORTÉS, A., PEREZ-MURCIA, M.D., GÓMEZ, I.: Co phytoavailability for tomato in amended calcareous soils. Bioresour. Technol., 96, 2005, 649-655.
- PILON-SMITS, E.A.H.: Phytoremediation. Ann. Rev. Plant Biol., 56, 2005, 15-39.
- RASKIN, I., KUMAR, P.B.A.N., DUSHENKOV, S., SALT, D.E.: Bioconcentration of heavy metals by plants. Curr. Opin. Biotechnol., 5, 1994, 285-290.
- SALT, D.E., PRINCE, R.C., BAKER, A.J.M., RASKIN, I., PICKERING, I.J.: Zinc ligands in the metal hyperaccumulator *Thlaspi caerulescens* as determined using X-ray absorption spectroscopy. Environ. Sci. Technol., 33, 1999, 713-717.
- SOUDEK, P., TYKVA, R., VANKOVA, R., VANEK, T.: Accumulation of radioiodine from aqueous-solution by sunflower (*Helianthus annuus* L.) at the laboratory scale. Environ. Exp. Bot., 57(3), 2006, 220-225.
- SURESH, B., RAVISHANKAR, G.A.: Phytoremediation A novel and promising approach for environmental clean-up. Crit. Rev. Biotechnol., 24(2-3), 2004, 97-124
- VRTOCH, L., HORNIK, M., PIPISKA, M., AUGUSTIN, J.: Radiocesium bioaccumulation by hydroponically cultivated tobacco plant (*Nicotiana tabacum* L.). Nova Biotechnol., VI-I, 2006, 19-28.
- VRTOCH, L., PIPISKA, M., HORNIK, M., AUGUSTIN, J.: Bioaccumulation of $^{60}\text{Co}^{2^+}$ ions in tobacco plants. Nova Biotechnol., VII-I, 2007, 41-49.
- WHITE, P.J., BROADLEY, M.R.: Mechanisms of caesium uptake by plants. New Phytol., 147, 2000, 241-256.
- ZHU, Y.G., SHAW, G.: Soil contamination with radionuclides and potential remediation. Chemosphere, 41, 2000, 121-128.